

**GENOTYPING BY PCR PROTOCOL**  
**MUTANT MOUSE RESOURCE & RESEARCH CENTER: UC DAVIS**

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530-754-MMRRC

Protocol Name: B6J.Cg-Tg(Tmc1-Tmc1\*)1Ajj/Mmucl MMRRC: 050609-UCD

Protocol:

Reagent/Constituent	Volume ( $\mu$ L)
Water	5.6
GoTaq® G2 Colorless Master Mix, 2X	7.5
Primer 1. (stock concentration is 20 $\mu$ M)	0.45
Primer 2. (stock concentration is 20 $\mu$ M)	0.45
DNA (example) extracted w/ "Qiagen DNeasy columns or other similar silica based kits"	1.0
	<b>TOTAL VOLUME</b>
	15

**Comments on protocol:**

- Protocol may work with other DNA extraction methods.
- Use Touch-Down cycling protocol-first 10 cycles anneal at 65°C decreasing in temperature by 1.0°C; next 30 cycles anneal at 55°C.
- PCR shown is form 50608 because 50609 DNA was not available.
- Primers 3 through 6 are for the background strain JAX#09146. Primers 5 & 6 are generic LacZ primers.

**Strategy:**

Steps	Temp (°C )	Time (m:ss)	# of Cycles
1. Initiation/Melting HOT START? <input type="checkbox"/>	94	5:00	<b>1x</b>
2. Denaturation	94	0:15	
3. Annealing steps 2-3-4 cycle in sequence	65 ( $\downarrow 1^{\circ}\text{C}/\text{cycle}$ )	0:30	<b>10x</b>
4. Elongation	72	0:40	
5. Denaturation	94	0:15	
6. Annealing steps 5-6-7 cycle in sequence	55	0:30	<b>30X</b>
7. Elongation	72	0:40	

**Primers:**

**Electrophoresis Protocol:**

Name	Nucleotide Sequence (5' - 3')	Agarose: 1.5%	V: 90
1. 50609-F	GGCCGCTAAATACGACTCACTAT	Estimated Running Time: 90 min.	
2. 50609-R	AGTAGAGTCCAGGCATGCTAAAAT	Primer Combination	Band (bp)
3. 09146-wtF	GACAGTTGGTGCTGGGATCT	1 & 2	342
4. 09146-wtR	AAGTACGAGGCCACTGAGGA	3 & 4	215
5. 09146-LaczF	CGTGGCCTGATTCAATTCC	5 & 6	315
6. 09146-LaczR	ATCCTCTGCATGGTCAGGTC		

