

**GENOTYPING BY PCR PROTOCOL**  
**MUTANT MOUSE REGIONAL RESOURCE CENTER: UC DAVIS**

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 530-754-MMRRC

NAME OF PCR: C57BL/6J-Tlr6<sup>m2B<sup>tlr</sup></sup>/Mmucd, (m2sd1) MMRRC # 030959-UCD

**Protocol:**

Reagent/ Constituent	Volume (µL)
Water	20.0
10x Buffer (contains 15mM MgCl <sub>2</sub> )	2.5
dNTPs (stock concentration is 25mM)	0.5
Primer 1 (stock concentration is 20µM) PCR F	0.5
Primer 2 (stock concentration is 20µM) PCR R	0.5
Taq Polymerase	0.5
gDNA template (50-100ng/µl) extracted with <input type="checkbox"/> NaOH <input type="checkbox"/> Proteinase K <input checked="" type="checkbox"/> Other: Any	0.5
<b>TOTAL VOLUME OF REACTION:</b>	<b>25µL</b>

**Comments on protocol:**

- *M2sd1* genotyping is performed by amplifying the region containing the mutation using PCR, followed by sequencing of the amplified region to detect the single nucleotide change.
- lab uses JumpStart@REDTaq@ReadyMix@ (P1107- Sigma), 12.5 µl in a 25 µl reaction (includes Taq, buffer, dNTPs).
- Confirm the size and presence of the amplicon on a 1X TAE gel. Purify using spin dialysis or the QIAGEN QIAquick PCR purification kit. Sequence the purified DNA fragments utilizing the Sequencing Primers listed.

**Strategy:**

Steps	Temp (°C)	Time (m:ss)	# of Cycles
1. Initiation/Melting HOT START? <input checked="" type="checkbox"/>	94	2:00	1
2. Denaturation	94	0:30	} 30x
3. Annealing } steps 2-3-4 will cycle in sequence	56	0:30	
4. Elongation }	72	1:00	
5. Amplification	72	7:00	1
6. Finish	4	∞	n/a

**Primers:**

Name	Nucleotide Sequence (5' - 3')
1. M2sd1 PCR F	GGAGACAGCACTGAAGTCACTGATG
2. M2sd1 PCR R	TGGCACCCTCACTCTGGATGAAG
3. M2sd1 _Seq (F)	TTTCAAAGGAGGCGCTATACTCG

**Electrophoresis Protocol:**

Agarose: 2% mV: 80 Estimated Running Time: 90 min

Primer Combination	Band	Genotype
1 and 2	1.24 kB	<i>m2sd1</i>
<b>SNP found at position ~ 1501 of sequencing</b>		

Mutation site (red) and flanking sequence:

WT agagat **A**gcagac  
*M2sd1* agagat **T**gcagac