

**GENOTYPING BY PCR PROTOCOL**  
**MUTANT MOUSE REGIONAL RESOURCE CENTER: UC DAVIS**

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530-754-MMRRC

NAME OF PCR: B6.129S1-Syngap1<sup>tm1.2Mabk</sup>/Mmcd

MMRRC # 031831-UCD

**Protocol:**

Reagent/ Constituent	Volume (µL)
Water	10.775
10x Buffer (contains / without 15mM MgCl <sub>2</sub> )	2.5
MgCl <sub>2</sub> (stock concentration is 25mM)	1.7
Betaine (stock concentration is 5M) <i>Optional</i>	6.5
dNTPs (stock concentration is 10mM)	0.5
DMSO <i>Optional</i>	0.325
Primer 1 (stock concentration is 20µM)	0.5
Primer 2 (stock concentration is 20µM)	0.5
Primer 3 (stock concentration is 20µM)	0.5
Taq Polymerase 5Units/µL	0.2
DNA extracted with "Qiagen DNeasy columns or other similar silica based kits"	1.0
<b>TOTAL VOLUME OF REACTION:</b>	<b>25µL</b>

**Comments on protocol:**

- Protocol may work with other DNA extraction methods.
- Use Touch-Down cycling protocol-first 10 cycles anneal at 65° C decreasing in temperature by 1.0° C; next 30 cycles anneal at 55° C.
- Betaine and DMSO have been standardized due to high GC content. Protocol may be tested without. Also, may adjust MgCl<sub>2</sub> to increase reaction or decrease non specific amplifications.

**Strategy:**

Steps	Temp (°C)	Time (m:ss)	# of Cycles
1. Initiation/Melting <span style="float:right">HOT START? <input type="checkbox"/></span>	94	5:00	1
2. Denaturation	94	0:15	} 40x
3. Annealing } steps 2-3-4 will cycle in sequence	65 to 55 (↓1°C/cycle)	0:30	
4. Elongation	72	0:40	
5. Amplification	72	5:00	
6. Finish	4	∞	n/a

**Primers:**

Name	Nucleotide Sequence (5' - 3')
1: 031831-mutR	GAAGAGGAGTTTACGTCCAGCCAAGCT
2: 031831-wtR	CGGATGCTATGTGCAGTGGTGGGA
3: 031831-comF2	GAAGTACCAATGGCATCCTTGAGG

**Electrophoresis Protocol:**

Agarose: 1.3%      V: 90      Estimated Running Time: 90 min.

Primer Combination	Expected Bands	Genotype
1 and 3	243 bp	mutant -/-
2 and 3	380 bp	wild-type +/+