

GENOTYPING BY PCR PROTOCOL
MUTANT MOUSE REGIONAL RESOURCE CENTER: UC DAVIS
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 530-754-MMRRC

NAME OF PCR: C57BL/6J-Nlrp3^{m1Btlr}/Mmcd, (ND1) MMRRC # 032646-UCD

Protocol:

Reagent/ Constituent	Volume (μL)
Water	20.0
10x Buffer (contains 15mM MgCl ₂)	2.5
Betaine (stock concentration is 5M) <i>Optional</i>	6.5
dNTPs (stock concentration is 25mM)	0.5
DMSO <i>Optional</i>	0.325
Primer 1 (stock concentration is 20μM) ND1 PCR (F)	0.5
Primer 2 (stock concentration is 20μM) ND1 PCR (R)	0.5
Taq Polymerase	0.5
DNA sample extracted with <input type="checkbox"/> NaOH <input type="checkbox"/> Proteinase K <input checked="" type="checkbox"/> Other: Any	0.5
TOTAL VOLUME OF REACTION:	25μL

Comments on protocol:

- PCR products are verified to contain the correct amplicon size by running ~10μl of the reaction on a gel and the remaining 15μl purified via column based PCR purification method for sequencing.
- Use Touch-Down cycling protocol-first 10 cycles anneal at 65° C decreasing in temperature by 1.0° C; next 30 cycles anneal at 55° C.
- Betaine and DMSO have been standardized due to high GC content. Protocol may be tested without. Also, may adjust MgCl₂ to increase reaction or decrease non specific amplifications.

Strategy:

Steps	Temp (°C)	Time (m:ss)	# of Cycles
1. Initiation/Melting HOT START? <input type="checkbox"/>	94	5:00	1
2. Denaturation	94	0:15	} 10x
3. Annealing	65 to 55 (↓1°C/cycle)	0:30	
4. Elongation		0:40	
5. Denaturation	94	0:15	} 30x
6. Annealing	55	0:30	
7. Elongation		0:40	
8. Amplification	72	5:00	1
9. Finish	15	∞	n/a

Primers:

Name	Nucleotide Sequence (5' - 3')
1: ND1 PCR (F)	ACA GCT TAA AGG CTA AGC CCC TGC
2: ND1 PCR (R)	TTC CAC GCC TAC CAG GAA ATC TCG
3: ND1_seq (F)	GGT TGG CTT CGA ACT CAG AAA TC
4: ND1_seq (R)	CTA AGG CAC GTT TTG TTT CAC G

Electrophoresis Protocol:

Agarose: 2% mV: 80 Estimated Running Time: 90 min

Primer Combination	Band	Genotype
1 and 2	921 bp	ND1
SNP found at position ~ of sequencing		

Mutation site (red) and flanking sequence:

WT accctctgTgaggtgctgaa
 ND1 accctctgGgaggtgctgaa