# GENOTYPING BY PCR PROTOCOL KOMP Repository: UC DAVIS

Protocol: CR956 Gia10

Reagent/Constituent	Volume (µL)
Water	10.275
10x Buffer	2.5
MgCl <sub>2</sub> (stock concentration is 25mM)	1.7
Betaine (stock concentration is 5M) Optional	6.5
dNTPs (stock concentration is 10mM)	0.5
DMSO Optional	0.325
Primer 1. (stock concentration is 20µM)	0.5
Primer 2. (stock concentration is 20µM)	0.5
Primer 3. (stock concentration is 20µM)	0.5
Primer 4. (stock concentration is 20µM)	0.5
Taq Polymerase 5Units/μL	0.2
DNA (example) extracted w/ "Qiagen DNeasy columns or other similar silica based kits"	1.0
TOTAL VOLUME OF REACTION:	25.000 μL

### Comments on protocol:

- Protocol may work with other DNA extraction methods.
- Use Touch-Down cycling protocol-first 10 cycles anneal at 65°C decreasing in temperature by 1.0°C; next 30 cycles anneal at 55°C.
- Betaine and DMSO have been standardized due to high GC content. Protocol may be tested without. Also, may adjust MgCl<sub>2</sub> to increase reaction or decrease non-specific amplifications.

Strategy:

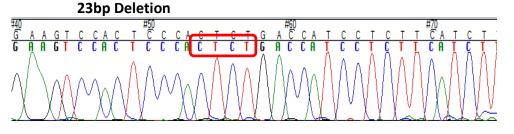
Steps		Temp (°C)	Time (m:ss)	# of Cycles
1. Initiation/Melting	HOT START? ☐	94	5:00	1
2. Denaturation		94	0:15	
3. Annealing	steps 2-3-4 cycle in sequence	65 to 55 (↓1°C/cycle)	0:30	40x
4. Elongation		72	0:40	
5. Amplification		72	5:00	1
6. Finish		15	$\infty$	n/a

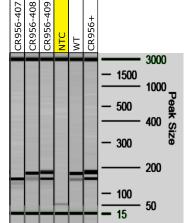
#### Primers:

#### **Electrophoresis Protocol:**

Name	Nucleotide Sequence (5' - 3')	Agarose: 1.5%	V: 90	
1. SEQ-Gja10-F	GCTTCTGTATGTTTCAGATGTGACTTCC	Estimated Running Time: 90 min		
2. SEQ-Gja10-R	GGACTGCTCATCATCCCAGACG	<b>Primer Combination</b>	Band (bp)	Genotype
		1 & 2	183	wildtype
		1 & 2	160	mutant

## Frameshift Indel Gja10 Exon 1





Contig gaagtccactccactccactatagtggggaagatctggctgaccatcctcttcatct

**Allele Description:** Exon 1 ENSMUSE00001407006 had 23 bp (ccactatagtggggaagatctgg) deleted from the Gja10 gene ENSMUST00000219644.1 using CRISPR Cas9 gene editing technology in mouse zygotes. This causes a frameshifted transcript followed by early termination signal. Subsequent founders were backcrossed to C57BL6/N to produce sequence confirmed heterozygous animals.