# GENOTYPING PROTOCOL MUTANT MOUSE RESOURCE \& RESEARCH CENTER: UC DAVIS 

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## Protocol Name: CR10818 Hacd1 exdel

Protocol:
GoTaq® G2 Colorless Master Mix(Promega)

| Reagent/Constituent | Volume $(\mu \mathrm{L})$ |
| :--- | :---: |
| Water | 4.5 |
| GoTaq® G2 Colorless Master Mix,2X | 7.5 |
| Primer 1. (stock concentration is $20 \mu \mathrm{M})$ comF | 0.5 |
| Primer 2. (stock concentration is $20 \mu \mathrm{M})$ wtR | 0.5 |
| Primer 3. (stock concentration is $20 \mu \mathrm{M})$ mutR | 0.5 |
| DNA (example) extracted w/ "Qiagen DNeasy columns or other similar silica based kits" | 1.5 |
|  | TOTAL VOLUME OF REACTION: |

## Comments on protocol:

- Protocol may work with other DNA extraction methods.


## Strategy:

| Steps | Temp ( $\left.{ }^{\circ} \mathrm{C}\right)$ | Time (m:ss) | \# of Cycles |
| :--- | :---: | :---: | :---: |
| 1. Initiation/Melting | 94 | $2: 00$ | 1x |
| 2. Denaturation START? | 94 | $0: 10$ |  |
| 3. Annealing | steps 2-3-4 cycle in sequence | $65\left(\downarrow 1^{\circ} \mathrm{C} / \mathrm{cycle}\right)$ | $0: 30$ |
| 4. Elongation | 68 | $2: 00$ | $\mathbf{1 0 x}$ |
| 5. Denaturation | 94 | $0: 15$ |  |
| 6. Annealing | 55 | $0: 30$ | $\mathbf{2 5 x}$ |
| 7. Elongation | 68 | $2: 00(\uparrow 20 \mathrm{sec} / \mathrm{cycle})$ |  |
| 8. Finish | 4 | $\infty$ | $\mathrm{n} / \mathrm{a}$ |

Primers:
Electrophoresis Protocol:

| Name | Nucleotide Sequence (5' - 3') | Agarose: 1.5\% $\quad$ V: $\mathbf{9 0}$ |  |  |
| :--- | :--- | :--- | :--- | :--- |
| 1. CR_Hacd1_comF | ATCTACAGTTTGTGTGACTGTCATCACC | Estimated Running Time: $\mathbf{9 0}$ min. |  |  |
| 2. CR_Hacd1_wtR | CGTACCATAGCAATAGCAAGAACCAA | Primer Combination | Band (bp) | Genotype |
| 3. CR_Hacd1_mutR | CCTTCTTCAGAGTCTCTGGTTCTGG | $1 \& 2,1 \& 3$ | 674,2283 | wildtype |
|  |  | $1 \& 3$ | 560 | mutant |

Allele Description: Exon 2 ENSMUSE00001337417, Exon 3 ENSMUSEO0001344764, Exon 4 ENSMUSEO0001343854 and flanking splicing regions were constitutively deleted from the Hacd1 gene ENSMUST00000091429.11 using CRISPR Cas9 gene editing technology in mouse zygotes. Subsequent founders were backcrossed to C57BL6/N to produce sequence confirmed heterozygous animals.
*wtR primer untested (ePCR verified)


